JOURNAL OF CLINICAL ONCOLOGY

# Targeting Cancer at the Nuclear Pore

Asfar S. Azmi and Ramzi M. Mohammad, *Wayne State University School of Medicine, Detroit, MI* See accompanying article on page 4142

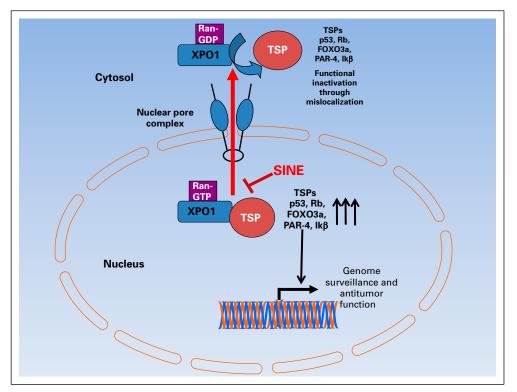
Nuclear protein transport regulates the function of cellular proteins in a compartmentalization-dependent fashion.<sup>1</sup> Proteins require an active form of transport through the nuclear pore that involves specialized transporters belonging to the karyopherin family (Fig 1).<sup>2</sup> The import of the majority of the cytosolic proteins is regulated by importins through nuclear localization signal recognition.<sup>3</sup> The export of nuclear proteins is almost exclusively mediated through exportin 1 (XPO1), also known as chromosome maintenance region 1 (CRM1), through the recognition of a conserved nuclear export signal (NES).<sup>2</sup> Nuclear export is a crucial aspect in the biology of any protein because mislocalization causes their functional inactivation. Most of the transcription factors and tumor suppressor proteins (TSPs), such as p53, IkB, p27, Rb, prostate apoptosis response-4 (PAR-4), and others, require nuclear retention and sequence-specific alignment to DNA to induce gene expression changes and transcriptional regulation of targets. It is logical to assume that the aberrant expression of nuclear transporters would result in unusual import or export of proteins, leading to their functional inactivation. Supporting this, there is ample evidence that cancer cells harbor an unusually higher expression of XPO1, making it an attractive therapeutic target to restore the proper localization of TSPs.<sup>4</sup>

XPO1/CRM1 can transport more than 200 target proteins.<sup>5</sup> Many of these cargoes are TSPs and transcription factors that can affect the majority of cancer hallmarks, driving the interest in the pharmaceutical industry as a target worth pursuing.<sup>5</sup> The first agent, leptomycin B (LMB), a natural product (originally developed as an antifungal compound), could specifically inhibit CRM1 export function.<sup>6</sup> LMB covalently binds to the NES recognizing Cys528 amino acid in XPO1/CRM1, thereby effectively blocking its target recognition.<sup>7</sup> Nevertheless, LMB showed numerous secondary effects and proved to be highly toxic and, therefore, was discontinued from a single phase I clinical trial.<sup>8</sup> Later, some modified forms of LMB (eg, leptomycin A) with better toxicity profiles were also tested preclinically.9 Ratjadone is another natural product that works as a CRM1 inhibitor in a fashion similar to LMB.<sup>10</sup> In parallel, CBS9106 was developed as a potent CRM1 inhibitor<sup>11</sup> with a mechanism distinct from that of LMB.<sup>12</sup>

Around 2010, a series of specific inhibitors of nuclear export (SINEs) developed at Karyopharm Therapeutics (KPT) in Natick, Massachusetts (now located in Newton, MA), were presented at the American Society of Hematology annual meeting. These agents were shown to effectively block nuclear export function by binding to the Cys528 amino acid in the NES-recognizing domain of XPO1. Unlike irreversibly binding LMB, the SINE compounds bind Cys528 in XPO1 in a slowly reversible fashion. Their specificity has been validated using CRISPR/Cas9 genome editing where a single mutation in Cys528 renders the drug ineffective.<sup>13</sup> These agents showed cancer cell selectivity and broad activity against a spectrum of solid tumors and hematologic cell lines as single agents. Numerous groups in heme models have demonstrated that SINE could synergize with chemotherapeutic agents such as cyclophosphamide, doxorubicin, vincristine, and prednisone; BRAF inhibitors; mammalian target of rapamycin inhibitors; and dexamethasone. Studies also showed synergy with gemcitabine and platinum compounds in solid tumor models (reviewed in Turner et al<sup>14</sup>). SINEs could also suppress cancer stemness and reverse epithelial-to-mesenchymal transition, which is an event considered critical to metastasis.<sup>15</sup> These multimodel analyses ushered the SINE compound selinexor into multiple phase I and II clinical trials.

The two recent articles published in Journal of Clinical Oncology (JCO) present the results of multi-institutional clinical studies on selinexor.<sup>16,17</sup> The first JCO article, by Abdul-Razak et al,<sup>16</sup> presents results from a multicenter phase I study that evaluated selinexor (KPT-330) and determined the recommended phase II dose. The study enrolled 189 patients with advanced solid tumors who received selinexor  $(3 \text{ to } 85 \text{ mg/m}^2)$ in either 3- or 4-week continuous cycles. The study used gene expression to evaluate pre- and post-treatment levels of XPO1 mRNA in patient-derived leukocytes.<sup>16</sup> The tumor biopsies were also examined by immunohistochemistry for changes in markers consistent with XPO1 inhibition. The main outcomes include the observation of dose-dependent elevations in XPO1 mRNA in leukocytes up to a dose level of 28 mg/m<sup>2</sup> before reaching a plateau. In line with the proposed mechanism of action of the drug, paired tumor biopsies showed nuclear accumulation of key TSPs, reduction of cell proliferation, and induction of apoptosis. Among 157 patients evaluable for response, one complete and six partial responses were observed (n = 7, 4%), with 27 patients (17%) achieving stable disease for  $\geq 4$  months.<sup>16</sup>

In the second *JCO* article, Gounder et al<sup>17</sup> evaluated pharmacodynamic changes in tumor biopsies of 54 patients treated with oral selinexor administered twice per week (administered on days 1 and 3) at one of three doses (30 mg/m<sup>2</sup>, 50 mg/m<sup>2</sup>, or a flat dose of 60 mg) either continuously or on a schedule of 3 weeks on and 1 week off. Pharmacokinetic analysis was performed under fasting and feeding states (low- v high-fat content) and using various formulations of selinexor (tablet, capsule, or suspension).<sup>17</sup> Most strikingly, there



**Fig 1.** Nuclear protein export is a druggable avenue in cancer. Proteins with size larger than 40 kDa are exported from the nucleus to the cytoplasm with the aid of specialized carrier protein export in 1 (XPO1), also known as chromosome maintenance region 1 (CRM1). XPO1 recognizes a nuclear export signal (NES) sequence in the target protein. NES is a short amino acid sequence of a few hydrophobic residues in the majority of proteins. XPO1 recognizes this NES, allowing attachment that results in movement of cargo protein from cell nucleus to the cytoplasm through the nuclear pore complex. Nuclear export of proteins occurs first through the binding of Ran-GTP (a G protein) to XPO1. Such binding causes structural alteration in XPO1, thereby increasing its affinity toward protein that is to be exported. Once the cargo protein is bound, the Ran-exportin-cargo complex moves out of the nucleus through the nuclear pore in a process that is controlled by the guanosine triphosphate (GTP)–guanosine diphosphate (GDP) concentration gradient. Once in the cytosl, the GTPase activating proteins (GAPs) then hydrolyze the Ran-GTP to Ran-GDP, and this causes structural alteration and subsequent XPO1 release. In the absence of Ran, the XPO1 molecule loss affinity for the nuclear cargo as well. Tumor suppressor proteins (TSPs) such as p53, forkhead box 03 (FOXO3a), prostate apoptosis response 4 (PAR-4), cyclin-dependent kinase inhibitor 1B (p27), retinoblastoma (Rb), and others are export (SINE) compounds block nuclear export by binding to the NES recognizing Cys528 in CRM1, resulting in nuclear accumulation of important genome surveillance transcription factors and TSPs.

was evaluable nuclear retention of TSPs in paired tumor biopsies alongside decreased cell proliferation, increased apoptosis, and stromal deposition.<sup>17</sup> Although none of the 52 patients evaluated showed objective response (Response Evaluation Criteria in Solid Tumors [RECIST] version 1.1), 33% of the patients showed stable disease. Similar to the findings of Abdul-Razak et al,<sup>16</sup> this study also reported drug-related grade 1 or 2 adverse events such as nausea, vomiting, anorexia, and fatigue, which were well managed with supportive care. The commonly reported grade 3 or 4 toxicities in this study were anemia, thrombocytopenia, fatigue, leukopenia, and lymphopenia. Selinexor was better tolerated when administered as a flat dose on an intermittent schedule. Encouragingly, clinically significant major organ or cumulative toxicities were rare. It is likely that the recent introduction of KPT-8602 (an analog of selinexor with better toxicity profile) into a phase I trial (ClinicalTrials.gov identifier: NCT02649790) is to specifically address and overcome these toxicity-related issues of the parent compound.<sup>18,19</sup>

These are encouraging findings and could form the basis for the expansion into phase II clinical studies. Such efforts are under way, and a number of phase IB/II studies are ongoing, including studies of selinexor, gemcitabine, and nanoparticle albumin-bound paclitaxel in metastatic pancreatic cancer (NCT02178436), gliomas (NCT01986348), and gynecologic and metastatic breast malignancies (NCT02025985); selinexor plus sorafenib combination in leukemia (NCT02530476); and selinexor plus cyclophosphamide, doxorubicin, vincristine, and prednisone in non-Hodgkin lymphoma (forthcoming). In parallel, the SINE analog KPT-335 (verdinexor) has been evaluated in a phase I study with clinical benefits and partial response in spontaneous canine non-Hodgkin lymphoma.<sup>20</sup>

Despite these encouraging results, the optimism is still mixed with caution. Some unwelcomed oncogenes are bound to be also retained in the nucleus, causing an unfavorable balance between TSPs and oncogenes. High-throughput proteomic approaches may help evaluate the consequence of such global nuclear retention of proteins in cancer and normal cells to identify the mechanism of selinexor's cancer cell selectivity. It would also be worthy to investigate the epigenetic changes induced by selinexor in vitro, in vivo, and in patients given the emerging knowledge that aside from proteins, the export of noncoding RNAs is also regulated by XPO1.<sup>21</sup> These approaches are currently under way in our laboratory and may provide clues to better management of toxicity-related issues.

In summary, the recent publication of phase I studies on selinexor has certainly marked a major turn in the field of nuclear export inhibitors. These studies will definitely bolster the expansion phase and perhaps clinical approval of selinexor and related analogs for the treatment of cancer at the nuclear pore.

#### Support

Supported by the National Institutes of Health Grant No. NIH 1R21CA188818-02 and also by the SKY Foundation, the James H. Thie Memorial Foundation, Karyopharm Therapeutics, and the Perri Foundation for Pancreatic Cancer.

### REFERENCES

1. Strambio-De-Castillia C, Niepel M, Rout MP: The nuclear pore complex: Bridging nuclear transport and gene regulation. Nat Rev Mol Cell Biol 11: 490-501, 2010

2. Fung HY, Chook YM: Atomic basis of CRM1cargo recognition, release and inhibition. Semin Cancer Biol 27:52-61, 2014

3. Chook YM, Blobel G: Karyopherins and nuclear import. Curr Opin Struct Biol 11:703-715, 2001

**4.** Mahipal A, Malafa M: Importins and exportins as therapeutic targets in cancer. Pharmacol Ther 164: 135-143, 2016

 Xu D, Marquis K, Pei J, et al: LocNES: A computational tool for locating classical NESs in CRM1 cargo proteins. Bioinformatics 31:1357-1365, 2015

6. Nishi K, Yoshida M, Fujiwara D, et al: Leptomycin B targets a regulatory cascade of crm1, a fission yeast nuclear protein, involved in control of higher order chromosome structure and gene expression. J Biol Chem 269:6320-6324, 1994

 Kudo N, Wolff B, Sekimoto T, et al: Leptomycin B inhibition of signal-mediated nuclear export by direct binding to CRM1. Exp Cell Res 242: 540-547, 1998

8. Newlands ES, Rustin GJ, Brampton MH: Phase I trial of elactocin. Br J Cancer 74:648-649, 1996

9. Mutka SC, Yang WQ, Dong SD, et al: Identification of nuclear export inhibitors with potent anticancer activity in vivo. Cancer Res 69:510-517, 2009

**10.** Köster M, Lykke-Andersen S, Elnakady YA, et al: Ratjadones inhibit nuclear export by blocking CRM1/exportin 1. Exp Cell Res 286:321-331, 2003

**11.** Sakakibara K, Saito N, Sato T, et al: CBS9106 is a novel reversible oral CRM1 inhibitor with CRM1 degrading activity. Blood 118:3922-3931, 2011

12. Saito N, Sakakibara K, Sato T, et al: CBS9106induced CRM1 degradation is mediated by cullin ring ligase activity and the neddylation pathway. Mol Cancer Ther 13:3013-3023, 2014

**13.** Neggers JE, Vercruysse T, Jacquemyn M, et al: Identifying drug-target selectivity of small-molecule CRM1/XPO1 inhibitors by CRISPR/Cas9 genome editing. Chem Biol 22:107-116, 2015

**14.** Turner JG, Dawson J, Cubitt CL, et al: Inhibition of CRM1-dependent nuclear export sensitizes malignant cells to cytotoxic and targeted agents. Semin Cancer Biol 27:62-73, 2014

**15.** Azmi AS, Muqbil I, Wu J, et al: Targeting the nuclear export protein XPO1/CRM1 reverses epithelial to mesenchymal transition. Sci Rep 5:16077, 2015

**16.** Abdul Razak AR, Mau-Soerensen M, Gabrail NY, et al: First-in-class, first-in-human phase I study of selinexor, a selective inhibitor of nuclear export, in patients with advanced solid tumors. J Clin Oncol [epub ahead of print on February 29, 2016]

## AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Disclosures provided by the authors are available with this article at www.jco.org.

# **AUTHOR CONTRIBUTIONS**

## Manuscript writing: All authors Final approval of manuscript: All authors

17. Gounder MM, Zer A, Tap WD, et al: Phase IB study of selinexor, a first-in-class inhibitor of nuclear export, in patients with advanced refractory bone or soft tissue sarcoma. J Clin Oncol 34:3166-3174, 2016

18. Hing ZA, Fung HY, Ranganathan P, et al: Nextgeneration XPO1 inhibitor shows improved efficacy and in vivo tolerability in hematological malignancies. Leukemia [epub ahead of print on June 21, 2016]

**19.** Etchin J, Berezovskaya A, Conway AS, et al: KPT-8602, a second-generation inhibitor of XPO1mediated nuclear export, is well tolerated and highly active against AML blasts and leukemia-initiating cells. Leukemia [epub ahead of print on June 24, 2016]

**20.** London CA, Bernabe LF, Barnard S, et al: Preclinical evaluation of the novel, orally bioavailable selective inhibitor of nuclear export (SINE) KPT-335 in spontaneous canine cancer: Results of a phase I study. PLoS One 9:e87585, 2014

**21.** Muqbil I, Bao B, Abou-Samra AB, et al: Nuclear export mediated regulation of microRNAs: Potential target for drug intervention. Curr Drug Targets 14: 1094-1100, 2013

DOI: 10.1200/JCO.2016.67.5637; published online ahead of print at www.jco.org on October 26, 2016.

# **AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST**

### Targeting Cancer at the Nuclear Pore

The following represents disclosure information provided by authors of this manuscript. All relationships are considered compensated. Relationships are self-held unless noted. I = Immediate Family Member, Inst = My Institution. Relationships may not relate to the subject matter of this manuscript. For more information about ASCO's conflict of interest policy, please refer to www.asco.org/rwc or jco.ascopubs.org/site/ifc.

Asfar S. Azmi

Research Funding: Karyopharm Therapeutics

Ramzi M. Mohammad Research Funding: Karyopharm Therapeutics